Bovine endometrial cells do not mount an inflammatory response to Leptospira

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Abstract

Leptospirosis causes abortion, premature birth, and stillbirth in cattle, but the mechanisms remain unclear. Infected cattle shed Leptospira intermittently and present a range of clinical symptoms, making diagnosis difficult. The primary route of Leptospira transmission in any animal is the colonization of the renal tubule and excretion by urine; however, Leptospira can also colonize the female reproductive tract of cows and can be transmitted by semen. Vaccination against Leptospira in the US is routine in cattle, but immunity is not guaranteed. The cell wall of Leptospira contains toll-like receptor agonists including peptidoglycan and lipopolysaccharide. The capacity of Leptospira to initiate an innate inflammatory response from uterine endometrial cells is unknown but may be a cause of reproductive failure. Using cell culture, we tested the capacity of bovine endometrial epithelial cells or human monocytes to elicit an inflammatory response to Leptospira borgpetersenii serovar Hardjo strain TC273. Cells were exposed to either heat-killed Leptospira, Leptospira outer membrane, Escherichia coli lipopolysaccharide, Pam3CSK4 or medium alone for 2 to 24 h. Exposure of bovine endometrial epithelial cells or human monocytes to heat-killed Leptospira or Leptospira outer membrane did not induce the expression of IL1A, IL1B, IL6, or CXCL8, while exposure to E. coli lipopolysaccharide or Pam3CSK4 increased the expression of IL1A, IL1B, IL6, and CXCL8 compared to control cells. This data suggest that Leptospira does not trigger a classical inflammatory response in endometrial cells. Understanding the interaction between Leptospira and the female reproductive tract is important in determining the mechanisms of Leptospirosis associated with reproductive failure.

Lay summary

Cows infected with the Leptospira have abortion and stillbirth. It is not known how Leptospira causes pregnancy failure in the cow. We tested if Leptospira causes inflammation in cells of the uterus which triggers pregnancy failure. We collected cells from the uterus of healthy cows at the abattoir and placed them into culture with Leptospira and measured the expression of genes associated with inflammation. To our surprise, cells of the uterus did not respond to Leptospira; however, the same cells did respond to other disease-causing bacteria found in the uterus. This suggests that cells of the uterus can recognize bacteria and produce an inflammatory reaction but not in response to Leptospira. This finding suggests the immune system of the uterus cannot detect Leptospira which may go on to cause reproductive failure in cows. Understanding how Leptospira interact with cells of the uterus will help reduce pregnancy failure of cows with leptospirosis.

Key Words: ▶ leptospirosis ▶ cattle ▶ reproduction ▶ inflammation ▶ endometrium

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Introduction

Leptospirosis is a worldwide bacterial zoonotic disease caused by pathogenic species of Leptospira. Leptospirosis affects over 1 million people annually, resulting in 58,900 deaths (Costa et al. 2015, Torgerson et al. 2015). Numerous mammalian species serve as hosts for Leptospira including rodents and cattle (Nally et al. 2016). Leptospira infection occurs after exposure to environments contaminated by spirochetes, and the subsequent colonization of the renal tubule that results in intermittent excretion of spirochetes in the urine and further contamination of the environment. Infected hosts may be asymptomatic or present a variety of clinical symptoms including fever, liver failure, respiratory distress, and reproductive failure (Ellis 2015).

In cattle, the greatest impact of leptospirosis is abortion, stillbirth, premature birth, reproductive failure, and milk drop syndrome (Ellis 2015, Loureiro & Lilenbaum 2020). Cattle are susceptible to infection with multiple Leptospira species and serovars including L. borgpetersenii serovar Hardjo, L. interrogans serovar Pomona, L. kirschneri serovar Grippotyphosa, and L. noguchii (Miller et al. 1991, Martins et al. 2015, Nally et al. 2018). The most prominent species of Leptospira associated with reproductive failure in cattle is L. borgpetersenii serovar Hardjo which decreases conception rate and increases calving to conception interval (Ellis et al. 1981, Miller et al. 1991, Guitian et al. 1999, Rajeev et al. 2014). Vaccination of cattle to serovars Canicola, Grippotyphosa, Hardjo, Icterohaemorrhagiae, and Pomona is common in the US and is an effective measure to prevent losses due to abortion and milk production. However, vaccination does not prevent infection and renal colonization, which results in vaccinated animals becoming carriers of Leptospira (Hanson et al. 1972, Srivastava 2006). In addition, host responses vary following vaccination suggesting a lack of complete and long-lasting protection, especially to L. borgpetersenii serovar Hardjo (Bolin et al. 1989, 1991, Rodrigues et al. 2011). A large proportion of the United States cattle herd is seropositive for pathogenic Leptospira (up to 49%), while seronegative cows can still excrete spirochete to transmit the infection to other animals (Miller et al. 1991, Talpada et al. 2003, Nally et al. 2018).

In cattle, leptospires have been isolated from the oviduct, uterus, aborted fetuses, and follicular fluid (Bienalski & Surujballi 1998, Bienalski et al. 1998, Monte et al. 2015, Loureiro & Lilenbaum 2020). Leptospires can migrate to the upper reproductive tract when administered intranasally or intracervically (Bienalski & Surujballi 1998, Bienalski et al. 1998). Interestingly, the detection of Leptospira in vaginal mucus by PCR is poorly correlated with detection in urine (Loureiro et al. 2017, Pinna et al. 2018). While leptospirosis causes reproductive failure in cattle, the mechanism by which this occurs is yet to be determined. It has been suggested that endometrial inflammation caused by Leptospira can change the developmental environment of the early conceptus, rendering the uterus hostile to pregnancy and resulting in reproductive failure (Loureiro & Lilenbaum 2020).

Postpartum uterine infection caused by gram-negative and gram-positive bacteria reduces reproductive success and causes localized inflammation of the endometrium in cattle. Epithelial and stromal cells of the endometrium respond to bacterial cell wall components, including lipopolysaccharide (LPS), via the toll-like receptor family and increase the expression of pro-inflammatory mediators including IL1B, IL6, and CXCL8 (Cronin et al. 2012, Turner et al. 2014). We hypothesized that bovine endometrial cells elicit an innate immune response to L. borgpetersenii serovar Hardjo. In addition, we aimed to determine the rate of Leptospira infection of cows by sampling urine, blood, and the uterus of vaccinated cows. Understanding the endometrial response to Leptospira infection will increase our knowledge of how reproductive failure occurs in cows with leptospirosis.

Materials and methods

Antigen preparation and immunoblotting

Leptospira borgpetersenii serovar Hardjo strain TC273 was isolated from a bovine urine sample in Iowa, as previously described (Nally et al. 2018). The virulence of TC273 was evaluated by intraperitoneal injection into Syrian hamsters (Mesocricetus auratus) as described by Nally et al. (2018). Outer-membrane fractions of low-passage virulent Leptospira borgpetersenii serovar Hardjo strain TC273 were enriched using Triton-X114 as previously described (Nally et al. 2001). Outer membrane (OM)-enriched fractions were compared to heat-killed leptospires by 1-D gel electrophoresis as previously described (Monahan et al. 2008). Total proteins were visualized by staining with Sypro Ruby (Invitrogen), and lipopolysaccharide was visualized by staining with Pro-Q Emerald 300 (Invitrogen) as per the manufacturer’s guidelines. For immunoblotting, samples were transferred to Immobilon-P transfer membrane (Millipore) and blocked overnight at 4°C with StartingBlock (TBS) blocking buffer (Thermo Fisher Scientific). Membranes were individually incubated with indicated antisera (anti-LipL21, anti-LipL32, and...
anti-LipL41 at 1:2500, 1:4000, and 1:2500, respectively or anti-Hardjo at 1:2500) in PBST for 1 h at room temperature, followed by incubation with horseradish-peroxidase anti-rabbit immunoglobulin G conjugate (Sigma–Aldrich). Bound conjugates were detected using Clarity Western ECL substrate (Bio-Rad Laboratories), and images were acquired using a Bio-Rad ChemiDoc MP imaging system.

**Bovine endometrial epithelial cell culture**

Bovine endometrial epithelial (BEND) cells were obtained from the American Type Culture Collection (ATCC, Manassas, VA; CRL-2398). Cells were cultured in complete culture medium (40% Ham F-12, 40% MEM, 10% fetal bovine serum, 10% horse serum, 100 IU/mL penicillin, 100 μg/mL streptomycin, 1.5 g/L of sodium bicarbonate, 0.034 mg/mL D-valine; Thermo Fisher Scientific) in 75 cm² flasks (Greiner Bio-One, Monroe, NC) at 38.5°C in a humidified atmosphere containing 95% air and 5% CO₂ until subconfluent. Cells were seeded on 24-well plates (TPP, Trasadingen, Switzerland) at a final density of 10⁵ cells/well in 500 μL and equilibrated for 24 h before the addition of treatments. Each experiment was performed seven times, with each replicate utilizing BEND cells between passages 3 and 13.

**Human THP-1 cell culture**

The human monocyte cell line, THP-1 (ATCC; TIB202) was used as a positive control for all treatments. THP-1 cells are a specialized immune cell line that elicits a strong innate immune response to pathogen-associated molecules, including *Escherichia coli*-derived LPS. Cells were cultured in 75 cm² flasks (Greiner Bio-One) in a complete culture medium containing RPMI 1640, 10% fetal calf serum, and 0.05 mM 2-mercaptoethanol (Thermo Fisher Scientific) at 37°C in a humidified atmosphere containing 95% air and 5% CO₂ until subconfluent. Cells were plated in 24-well plates (TPP) at a density of 10⁵ cells/mL in 500 μL. After 48 h, 50 ng/mL phorbol myristate acetate was added to promote the differentiation of monocytes to macrophage-like cells. Treatments were applied to THP-1 cells after differentiation. Each experiment was performed four times, with each replicate utilizing THP-1 cells between passages 12 and 14.

**Treatment of cultured cells**

Adherent THP-1 and BEND cells were exposed for 24 h to either ultrapure *E. coli* O111:B4 LPS (Invivogen), Pam3CSK4 (Invivogen), heat-killed *Leptospira borgpetersenii* serovar Hardjo (HK-lepto), *Leptospira borgpetersenii* serovar Hardjo outer membrane preparation (OM-lepto) or medium alone as a control. Doses of each treatment started at 10,000 ng/mL and decreased ten-fold to 1 ng/mL (maximal HK-lepto and OM-lepto treatments were 1000 ng/mL). To determine acute cellular responses, THP-1 and BEND cells were exposed to 100 ng/mL of LPS, Pam3CSK4, HK-lepto, OM-lepto or control medium for 2 or 12 h. Cell-free supernatants were collected, and total cellular RNA was stabilized in 350 μL of RLT buffer (Qiagen) and stored at −80°C.

**Evaluation of endometrial cell viability**

Viability of BEND cells was assessed by the cellular reduction of MTT (Thermo Fisher Scientific) as previously described (Rizo et al. 2019). Briefly, BEND cells were cultured in 96-well culture plates (TPP) at a density of 10⁵ cells/mL in 200 μL and equilibrated for 24 h at 38.5°C in a humidified atmosphere containing 95% air and 5% CO₂. Duplicate wells were exposed to 100 ng/mL of LPS, Pam3CSK4, HK-lepto, OM-lepto, or control medium for 2 or 24 h. Following treatment, 10 μL of MTT (5 mg/mL) was added to each well and incubated at 38.5°C for 4 h. Cells were washed with warm DPBS and lysed in 100 μL of dimethyl sulfoxide. Optical density of each well was measured using a microplate reader at 540 nm (BioTek). The blank corrected value for each well was determined, and an average optical density for each replicate was calculated using the average of duplicate wells. Data were normalized and expressed as fold change from cells treated with control culture medium only.

**RNA extraction and real-time RT-PCR**

Total RNA was extracted using the RNeasy Mini kit (Qiagen) according to the manufacturer’s instructions. Samples were quantified and checked for RNA quality by spectrophotometry (Nanodrop ND1000, Thermo Fisher Scientific) and then reverse-transcribed using the Verso cDNA kit according to the manufacturer’s instructions (Thermo Fisher Scientific). Primers were designed using the NCBI primer-design tool and verified by BLAST with the exception of human IL1A that was obtained from Sharkey et al. (2012) (Table 1). All primers were validated for amplification efficiency prior to sample analysis and conformed to MIQE guidelines (Pearson correlation coefficient r² > 0.98 and efficiency between 90 and 110%) (Bustin et al. 2009). Real-time PCR was performed in
20 µL reactions, containing 18 µL of SYBR Green Master Mix (Bio-Rad Laboratories) with 500 nM of each reverse and forward primer and 2 µL of template cDNA. The cycling was performed on a CFX Connect Real-Time PCR System (Bio-Rad Laboratories) consisting of an initial denaturation/ enzyme activation step for 30 s at 95°C followed by 39 PCR cycles using 5 s denaturation at 95°C, 30 s annealing at 60°C, and 10 s of extension at 65°C. Each reaction was performed in duplicate. A no-template control with no cDNA was included for each primer set to demonstrate the absence of non-specific amplification. Relative expression for genes of interest was calculated using the 2^ΔCt method relative to GAPDH for BEND cells and ACTB for THP-1 cells (Livak & Schmittgen 2001). Expression of GAPDH and ACTB was stable (P > 0.05) across all treatments (Supplementary Fig. 1, see section on supplementary materials given at the end of this article).

Tissue and fluid collection to determine the prevalence of Leptospira spp in blood, urine, and the uterus

The University of Florida Institutional Animal Care and Use Committee approved all animal procedures that were conducted at the University of Florida Dairy Research Unit. Cohort 2 consisted of 23 2-year old primiparous non-lactating Holstein cows born and raised in South Georgia and housed at the University of Florida Dairy Research Unit after calving.

Urine samples were collected from all 56 cows in cohorts 1 and 2 to determine the presence of Leptospira spp in the urinary tract by fluorescent antibody test (FAT) and culture (see below for details). Approximately 30 mL of midstream urine was collected from each cow into a sterile 50 mL conical tube after stimulation of the perineal area. Urine samples were immediately placed on ice and processed for live culture within 1 h of collection by adding two to three drops of fresh urine to individual culture tubes containing culture medium as previously described by Nally et al. (2018). The remaining urine was utilized for FAT analysis. Postmortem urine was collected from one cow in cohort 2, 63 days after the first urine collection. Within 30 min of slaughter, a sterile needle and syringe were used to aspirate urine from the bladder which was transferred to a sterile vial. The vial was maintained on ice for detection of Leptospira spp by FAT and live culture.

Whole blood was collected from the coccygeal vessel into evacuated tubes (Vacutainer, Becton Dickson, Franklin Lakes, NJ) without anticoagulant from six cows in cohort 2 prior to slaughter. Blood was collected 63 days after initial urine sampling. Whole blood was centrifuged for 10 min at 2400 × g at room temperature, and subsequent serum was frozen at −20°C. Serum samples were used to evaluate the exposure of cows to Leptospira antigen using the microscopic agglutination test (MAT; see below for details).

Uterine samples from cows in cohorts 1 and 2 were collected using the cytobrush technique immediately after initial urine collection as previously described (Rizo et al. 2019). Briefly, external genitalia were cleaned using 70% ethanol, and the cytobrush tool (Medscan Medical, Cooper Surgical, Trumball, CT) contained inside a metal sheath...
and covered by a sanitary chemise (WTA, Cravinhos, Brazil) was introduced into the vagina. Using rectal palpation, the tool was passed through the cervix and the sanitary chemise was retracted over the metal sheath. The cytobrush was then extended and placed in direct contact with the endometrium. The cytobrush was then rotated three times to sample endometrial cells and uterine fluid. The cytobrush was then retracted into the metal sheath and removed from the reproductive tract. Endometrial smears were prepared by gently rolling the cytobrush over a clean, glass slide (Thermo Fisher Scientific) and air-dried. Uterine samples were collected from a total of 27 cows and used for detection of *Leptospira* by FAT.

**Fluorescence antibody test to determine the presence of Leptospira spp**

The FAT was performed as previously described (Nally et al. 2018). Briefly, urine samples were centrifuged at 10,000 g for 30 min at 4°C. The resultant cell pellet was resuspended in 2 mL H$_2$O and 1 mL was transferred to a clean 1.5 mL tube and washed by centrifugation at 12,000 g for 10 min at 4°C. Approximately 100 μL of material was retained after centrifugation and was diluted in 500 μL of H$_2$O and centrifuged at 12,000 g for 10 min at 4°C. The supernatant was removed until approximately 50 μL remained which was then resuspended. A 15 μL aliquot of the resultant suspension was deposited on a 7 mm well FAT glass slide in duplicate. Slides were air-dried and fixed in acetone for 10 min and placed in a humidified chamber. Each well was incubated at 37°C for 1 h with 20 μL of high-titer rabbit anti- *Leptospira* sera (National Veterinary Services Laboratories, APHIS, USDA, Ames, IA) conjugated to fluorescein isothiocyanate. Slides were washed in PBS with gentle rocking for 10 min, dried and counterstained with Flazo Orange (National Veterinary Services Laboratory), followed by a final wash in PBS before coverslips were mounted using Vectashield (Vector Laboratories, Burlingame, CA). Microscopic examination was performed using a Nikon Eclipse E800 microscope and B2-A filter (excitation, 450–490 nm; emission, 520 nm) at 200× magnification. Positive FAT samples were categorized by the fluorescein isothiocyanate fluorescence that conformed to *Leptospira* morphology.

**Microscopic agglutination test**

Serum of six cows in cohort 2 was evaluated for the presence of antibodies to six live *Leptospira* serovars; *L. interrogans* serogroup Australis serovar Bratislava strain Jez Bratislava, *L. interrogans* serogroup Canicola serovar Portlandvere strain 12-001, *L. kirschneri* serogroup Grippotyphosa serovar Grippotyphosa strain GR-01-082, *L. interrogans* serogroup Sejroe serovar Hardjo strain Hardjoprajitno, *L. interrogans* serogroup Icterohemorrhagiae serovar Copenhageni strain IC-02-001, and *L. interrogans* serogroup Pomona serovar Pomona strain Pomona. Briefly, sample serum was added to live *Leptospira* cultures at two-fold increasing dilutions beginning at 1:25, up to 1:800.

**Statistical analysis**

Data were analyzed using SPSS software V24.0 (IBM Analytics). Data were assessed for normality with Shapiro–Wilk test and log transformed when appropriate (described in figure legends). Data were analyzed using the generalized linear mixed model with dose, treatment, and time as fixed factors. Pairwise comparisons were performed between each dose and vehicle controls. Statistical significance was declared when $P \leq 0.05$.

**Results**

**Characterization of Leptospira preparations used for cell treatments**

Heat-killed *Leptospira borgpetersenii* serovar Hardjo (HK-lepto) and *L. borgpetersenii* serovar Hardjo outer membrane preparation (OM-lepto) used to treat BEND cell and THP-1 cells were evaluated for the presence of *Leptospira* specific proteins and LPS after completion of cell stimulation experiments to confirm bioactivity (Fig. 1). Both preparations of HK-lepto and OM-lepto showed the presence of LipL32 (32kDa; Fig. 1B), LipL41 (41 kDa; Fig. 1C), LipL21 (21kDa; Fig. 1D), and reactivity to anti-Hardjo antiserum (Fig. 1E). Both preparations of HK-lepto and OM-lepto showed the presence of LPS using the Pro-Q Emerald 300 LPS stain (Fig. 1F), in an expected different conformation than *E. coli* 055: B55 LPS.

**Effect of bacterial components in endometrial cell viability**

Exposure of BEND cells to LPS, Pam3CSK4, HK-lepto or OM-lepto for 2 h had no effect on cell viability compared to controls (Fig. 2). While exposure of BEND cells to Pam3CSK4, HK-lepto, or OM-lepto for 24 h did not affect cell viability, exposure to LPS for 24 h increased cell viability compared to medium only controls ($P < 0.05$).
Effect of bacterial components on the expression of inflammatory mediators in cultured cells

Exposure of BEND cells to 100, 1000, or 10,000 ng/mL of LPS for 24 h increased the expression of IL6 and CXCL8 compared to medium alone controls (Fig. 3C and D; P < 0.05). Exposure to 100, 1000, or 10,000 ng/mL of Pam3CSK for 24 h increased BEND cell expression of IL1B and CXCL8 compared to medium alone controls, while 1000 or 10,000 ng/mL of Pam3CSK4 increased the expression of IL6 compared to medium alone controls and exposure to 10,000 ng/mL of Pam3CSK4 increased the expression of IL1A compared to medium alone controls (Fig. 3E, F, G and H; P < 0.05). Exposure of BEND cells to either HK-lepto or OM-lepto did not alter the expression of IL1A, IL1B, IL6, or CXCL8 compared to medium alone controls (Fig. 3I, J, K, L, M, N, O and P).

Acute exposure of BEND cells to 100 ng/mL of LPS or Pam3CSK for 2 or 12 h increased the expression of IL1A, IL1B, IL6, and CXCL8 compared to medium alone controls (Fig. 4A, B, C, D, E, F, G and H; P < 0.05). However, exposure of BEND cells to either HK-lepto or OM-lepto for 2 or 12 h did not alter the expression of IL1A, IL1B, IL6, or CXCL8 compared to medium alone controls (Fig. 4A, B, C, D, E, F, G and H).

Exposure of THP-1 cells to 10, 100, 1000, or 10,000 ng/mL of LPS for 24 h increased the expression of IL1B and CXCL8 compared to medium alone controls, while exposure to 100, 1000, and 10,000 ng/mL of LPS for 24 h increased the THP-1 expression of IL1A and IL6 (Fig. 5A, B, C and D; P < 0.05). Exposure of THP-1 cells to 10, 100, 1000, or 10,000 ng/mL of Pam3CSK for 24 h increased the expression of IL1A, IL1B, and CXCL8 compared to medium alone controls, while exposure to 10, 100, 1000, or 10,000 ng/mL of Pam3CSK for 24 h increased the expression of IL1A, IL1B, and CXCL8 compared to medium alone controls, while exposure to 10, 100, 1000, or 10,000 ng/mL of Pam3CSK for 24 h increased the expression of IL1A, IL1B, and CXCL8 compared to medium alone controls, while exposure to 10, 100, 1000, or 10,000 ng/mL of Pam3CSK for 24 h increased the expression of IL1A, IL1B, and CXCL8 compared to medium alone controls.
ng/mL of Pam3CSK for 24 h increased the THP-1 cell expression of IL6 compared to medium alone controls (Fig. 5E, F, G and H; \( P < 0.05 \)). Exposure of THP-1 cells to either HK-lepto or OM-lepto for 24 h did not alter the expression of IL1A, IL1B, IL6, or CXCL8 compared to medium alone controls (Fig. SI, J, K, L, M, N, O and P).

All uterine samples collected from 27 cows by cytobrush (including the urine FAT-positive cow) were FAT negative.

Detection of *Leptospira* in urine and uterine samples

Urine collected by micturition from 56 cows was analyzed for the presence of *Leptospira* spp using FAT. Urine of one cow (1.8%) in cohort 2 was FAT-positive that showed a *Leptospira* morphology (Fig. 6). Postmortem urine from the bladder of this single positive cow was collected 63 days after the first urine collection and was FAT-negative. All urine samples were culture negative.

Microscopic agglutination test of serum

Serum of six cows in cohort 2 was used to determine the presence of *Leptospira* antibodies 63 days after the first urine FAT analysis (Table 2). Blood from the single FAT-positive cow (cow #1) was included in the evaluation. Of the six cows tested, three had a positive MAT of 1:25 or greater against one serovar, including Canicola and Grippotyphosa. The single urine FAT-positive cow was MAT positive for Canicola, while two FAT-negative cows were MAT positive for Grippotyphosa.
Discussion

Leptospirosis in cattle is common and causes several clinical complications including reproductive failure. While the mechanisms of leptospirosis-induced reproductive failure are unknown, spirochetes are located within the female reproductive tract and it has been proposed that uterine inflammation caused by *Leptospira* could result in reproductive failure (Loureiro & Lilenbaum 2020). Both gram-negative and gram-positive bacteria (and isolated components) induce TLR signaling in bovine endometrial cells that results in an innate immune response to pathogens with increased expression of inflammatory mediators including IL1A, IL1B, IL6, and CXCL8 (Cronin et al. 2012, Turner et al. 2014). Bovine endometrial epithelial and stromal cells express all ten TLRs and when activated by pathogens increase the expression of cytokines and chemokines to induce a cellular inflammatory response and clear the pathogenic agent (Davies et al. 2008, Cronin et al. 2012, Turner et al. 2014). Here, bovine endometrial epithelial cells and human monocytes increased the expression of IL1A, IL1B, IL6, and IL8 when exposed to ultrapure gram-negative LPS (TLR4 agonist) or the synthetic lipopeptide Pam3CSK4 (TLR2/1 agonist). However, when either cell type was exposed to heat-killed *Leptospira* or purified outer membrane preparations of *Leptospira* for 2, 12, or 24 h no change in IL1A, IL1B, IL6, and CXCL8 expression was observed. These data suggest that while bovine endometrial cells have the functional capacity to induce an inflammatory response to bacterial components, *Leptospira* either evade detection by endometrial epithelial cells or elicit a non-classical immune response that was not measured here.

Previous experimental data in female dogs and mares demonstrate that *Leptospira* infection induces endometrial inflammation and is associated with reproductive failure (Pinna et al. 2013, Wang et al. 2014). In parallel, exposure of bovine neutrophils to *Leptospira* induced minimal expression of inflammatory cytokines and slight neutrophil extracellular trap formation (Wilson-Welder et al. 2016). In addition, experimental inoculation of the...
utero with *Leptospira* increased pregnancy failure in cattle and increased *Leptospira* titers in blood and vaginal mucus (Dhaliwal *et al.* 1996). Collectively, these studies suggest that peripheral cells or endometrial cells elicit an inflammatory response to *Leptospira*; however, other studies conclude that intrauterine inoculation of *Leptospira* has no effect on fertility or clinical inflammation, suggesting secondary mechanisms which may be responsible for reproductive failure in cattle (Vahdat *et al.* 1983). Indeed, *Leptospira* pathogens have been isolated from aborted fetuses and embryos of infected dams and as such may directly impact the reproductive success of the host by directly targeting the conceptus as opposed to the female reproductive tract (Ellis *et al.* 1981, Bielanski *et al.* 1998).

Most recently, molecular mechanisms have been described by which *Leptospira* can evade detection of TLR4, TLR5, NOD1, and NOD2 of the innate immune system (Werts *et al.* 2001, Ratet *et al.* 2017, Holzapfel *et al.* 2020). Our own characterization of LPS shown here (Fig. 1), clearly indicates that *E. coli* and *Leptospira* LPS are structurally different and result in different molecular weights. Indeed, previous work demonstrates that *E. coli* LPS induces a strong immune response in rabbits compared to LPS purified from *Leptospira* (de Souza & Koury 1992). The *Leptospira* outer membrane preparation used here contains LPS, LipL32, LipL41, LipL21, glycolipids, lipoproteins, and various other pathogen-associated molecular patterns that could be involved in virulence and induce a host immune response. In humans, *L. interrogans* can evade recognition by TLR4 and activates macrophages using TLR2 (Werts *et al.* 2001). The process of *Leptospira* immune system evasion may be specific to the growth stage of the pathogen and the species of the host (Holzapfel *et al.* 2020). When cattle are infected with *Leptospira*, the absence of clinical symptoms is common despite the continued excretion of *Leptospira* in urine and the presence of the pathogen in the reproductive tract. It is suggested that persistent infection is due to specific host-pathogen mechanisms that dampen the host immune response to resident *Leptospira*. However, when excreted in urine *Leptospira* modify their protein and antigen expression and modulate post-translational protein modifications which may aid in pathogen evasion of the host immune system (Nally *et al.* 2005, 2011). Understanding the molecular mechanisms by which *Leptospira* evades recognition by the host immune system require careful investigation in target species using specific cell types, and likely *in vivo* experimentation.

Studies estimate that up to 49% of cattle in the US are seropositive for pathogenic *Leptospira*, and vaccination to serovar Hardjo can induce an inconsistent response that results in short-term immunization (Miller *et al.* 1991). Our results screening urine and uterine samples of immunized animals by FAT and culture did not identify any *Leptospira*-positive cattle at the University of Florida herd. In general, all cattle at the University of Florida dairy research unit are born and raised on-site, providing for a closed system. However, we also screened cattle that were imported from Georgia, with one cow providing a positive urine FAT that was later negative when urine was collected from the bladder at the time of slaughter. This cow demonstrated intermittent shedding of *Leptospira* in urine that has been observed previously (Cordonin *et al.* 2011) and may suggest the presence of other *Leptospira*-positive cows that were not detected during the time of sampling. In addition, no uterine samples tested by FAT or culture returned a positive result; however, there is a poor correlation between the detection of *Leptospira* in urine and the uterus (Loureiro *et al.* 2017). In parallel, the same cows were tested for *Leptospira* infection by MAT at least 200 days after immunization to serovars Canicola, Gryppotyphosa, Hardjo, Icterohaemorrhagiae, and Pomona. Considering the immunization status of the cows and inclusion of the single FAT-positive cow, MAT titers were surprisingly low or negative for tested cows. It is not clear from the very limited data presented here or from the available literature as to what MAT titers should be expected following *Leptospira* vaccination or *Leptospira* infection. It is important to note that the MAT assay is

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**Table 2** MAT results from sera collected from group 2.

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<th>Cow</th>
<th>Bratislava</th>
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<th>Icterohaemorrhagiae</th>
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1:25, agglutination of sample at 1:25 serum dilution; -, no agglutination.
based on immunoglobulin (Ig) M that tends to increase rapidly after vaccination but rapidly declines and may, therefore, provide an inaccurate measure of immune status (Negi et al. 1971). The cows surveyed in this study were vaccinated at least 200 days prior to sample collection, which may explain the low titers observed in the MAT. Bacterin vaccines, like the ones used in the cows surveyed here, can produce a negative MAT response while still protecting the animal from Leptospira infection, which can be confirmed by assays that measure IgG, such as hamster immunization assays. Our data suggest that Leptospira serovar Hardjo does not induce a classical inflammatory response in bovine endometrial epithelial cells or human monocytes based on the specific inflammatory markers used here (IL1A, IL1B, IL6, and CXCL8) under the specific conditions tested. Leptospira may possess specific molecular mechanisms to evade detection by the innate immune system or perhaps induce a non-classical response with induction of other cytokines and mediators not measured here. It is also important to highlight that our model utilized a cell line and not isolated primary cells, thus extrapolation to an active Leptospira infection in the cow can only be considered with caution.

In cattle, the endometrial epithelial layer is commonly lost at the time of parturition which exposes the underlying stroma to bacterial pathogens that commonly cause postpartum uterine disease. Our culture model only evaluated the effects of Leptospira on epithelial cells and did not account for the possible interaction of Leptospira components on the underlying stroma. Moreover, the two-dimensional culture system employed here does not replicate epithelial cell polarization observed in vivo, and bovine endometrial epithelial cells do exhibit a vectorial release of specific inflammatory mediators when challenged by pathogens (Healy et al. 2015). Indeed, the current study fails to evaluate the effects of live Leptospira on endometrial cells, which would likely kill endometrial cells and induce subsequent inflammation via release of damage-associated molecular patterns. Nonetheless, the present study supports previous findings on how Leptospira might be able to evade host response and shows that might be true also for the bovine reproductive tract. Further investigation is required to determine if this evasion from the immune system plays a role in Leptospirosis-induced reproductive failure in cattle.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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References
Davies D, Meade KG, Herath S, Eckersall PD, Gonzalez D, White JO, Conlan RS, O’farrelly C & Sheldon IM 2008 Toll-


Monte LG, Ridieri KF, Jorge S, Oliveira NR, Hartwig DD, Amaral MG, Hartleben CP & DellaGostin OA 2015 Immunological and molecular characterization of Leptospira interrogans isolated from a bovine foetus. Comparative Immunology, Microbiology and Infectious Diseases 40:41–45. (https://doi.org/10.1016/j.cimid.2015.04.001)


Srivastava SK 2006 Prospects of developing leptosomal vaccines for animals. Indian Journal of Medical Microbiology 24:331.


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